Fixing organoids for Histology Embedding organoids in low melting point agar

- If not already aliquoted, prepare 2% low melting point agar in water and put into 1.5mL snap cap tubes. Unused aliquots of 2% agar can be kept in the fridge for years
- 2. Warm a heating block to **65°C**. Heat the agar in the block until liquefied. In the meantime, prepare labelled snap cap tubes for all samples.
- 3. Once organoids are settled to the bottom of their tubes (in 70% EtOH), pipette excess ethanol away, leaving the organoids in approximately 100µL volume.
- 4. Transfer organoids in ethanol to corresponding labeled snap cap tubes.
- 5. QUICKLY add **300µL agar** per tube & mix carefully.
 - a. Avoid bubbles but mix thoroughly.
 - b. Work quickly as agar polymerizes/solidifies rapidly.
- 6. Centrifuge tubes for **5 minutes at 4°C at 500G** to aggregate the organoids and allow the agar to polymerize around them.
 - a. If unsatisfied w/ the organoids' location within the agar, put the snap cap tube back on the heating block for ~10 minutes and centrifuge again.
- 7. Label histology cassettes for each sample using **pencil**
- 8. Add small amount of ethanol to each tube (containing organoids + agar) and flick the tube very hard to dislodge the contents.
- 9. Remove agar w/ organoids embedded from tubes.
 - a. Gently grab the agar puck with forceps so as not to damage/break the agar; or
 - b. tip the whole agar puck out of the tube once it is dislodged.

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- 10. Make a **transverse** cut of the agar puck with bias towards the tip of the cone where the organoids are.
 - a. Do not cut the portion of the puck containing the organoids too thin or it could curl once embedded in paraffin, leading to them occupying different planes within the block.
- 11. Place both halves of the cut agar puck inside the labelled cassette for the sample. Add a **foam insert** to the cassette if you think the sample could fall out of the cassette.
- 12. Place the entire cassette into a bucket of 70% ethanol and submit for paraffin embedding.